

RETALIATION OF SALMONELLA

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SUMMARY

Typhoid fever is endemic in developing countries because of the non-availability of clean drinking water. Blood cultures on pyrexial patients were done. Of the 25 blood cultures, 24 yielded growth of *Salmonella typhi* and one paratyphi A. The isolates were sensitive to quinolones (100%), 3rd generation cephalosporins (95%), aztreonam (76%), amoxicillin and chloramphenicol (20%) and SMZ-TMP (0%). From eighty to hundred percent patients improved with quinolones, 50% with 13 chloramphenicol, 20% with Ceftriaxone and none with amoycillin. With the rising resistance of *Salmonella* to the currently available new toxic and expensive drugs we should adopt basic hygienic principles rather than spend time and money on drugs.

INTRODUCTION

There are nearly 2000 *Salmonella* serotypes most of which reside in animals, the exception being *Salmonella typhi* which invariably has human source.¹ These organisms cause enteric fevers which range from self-limited gastroenteritis to typhoid fever, a potentially life threatening illness. Typhoid fever results in high mortality and morbidity. It is water born and is endemic in developing countries where there is scarcity of potable water.¹

The R-factor-mediated multiple resistance noted since 1972-73 has influenced not only the cost of the treatment but also the toxicity of various newly administered drugs.² One of such single large size 98 MDa plasmid has been isolated. It encodes resistance to chloramphenicol, ampicillin, tetracycline, streptomycin, Sulphamethoxazole-Trimethoprim (SMZ-TMP) but shows sensitivity to naldixic acid, quinolones and 3rd generation cephalosporins.

MATERIAL AND METHODS

All those patients from OPD or casualty who were running a temperature of

101 F⁰ or above were admitted in Medical A ward of Postgraduate Medical Institute, Lady Reading Hospital, Peshawar, from January 1993 to January 1995. A proforma was prepared which included symptoms, signs and various investigations. Blood cultures were taken and only those positive for *Salmonella* were included in the study. Moreover, the drug responses were also recorded.

RESULTS

Most of the patients belonged to Peshawar and their ages ranged from 12 to 55 years. The minimum duration of hospital stay was three days and the maximum 16 days.

Blood culture yielded growth of *Salmonella typhi* in 24 patients and paratyphi A in only one. Sensitivity test showed that all 25 (100%) isolates were sensitive to quinolones, 24 (95%) to 3rd generation cephalosporins, 19 (75%) to aztreonam (Azactam), 15 (60%) to aminoglycosides, 13 (52%) to 1st generation cephalosporins, 2 (20%) each to amoxicillin and chloramphenicol and none to SMZ-TMP.

(Table-I) The sensitivity of Salmonella to various drugs in different antibiotic groups is shown in Table-II. Various quinolones were given to 21 patients and 20 of them improved. Ciprofloxacin was effective in all 8 patients (100%). Eight (80%) out of 10 patients responded to ofloxacin and 4 (80%) out of 5 patients responded to pefloxacin. The two patients who did not respond to ofloxacin and pefloxacin in 5 days, improved with ciprofloxacin and were included in that sub-group.

One patient clinically diagnosed as respiratory tract infection was started on cephazoline before culture was available and improved with the same antibiotic. Ceftriaxone was effective in only one (20%) of the five patients who received this drugs. Of the six patients who received Chloromycetin only three (50%) responded. None of the three patients responded to amoxicillin. SMZ-TMP and aztreonam were not used in this study.

DISCUSSION

In several parts of the world where typhoid fever is endemic there is serious concern about the emerging patterns of Salmonella species that are resistant to the currently and antibiotics.³ These organisms cause significant morbidity and mortality. Since the first report of its successful use in 1948, chloramphenicol has been the mainstay of therapy for long and is still the drug of first choice in many parts of the world i.e. India, China, USSR.^{4,7} More recent studies have shown SMZ-TMP, ampicillin and amoxicillin to be effective in cases caused by chloramphenicol-susceptible and chloramphenicol-resistant organisms. However, as has previously been noted, a number of cases due to chloramphenicol-resistant and ampicillin-resistant strains were seen in 1970 in Mexico and the United States and there were several deaths.⁵ Strains of Salmonella typhi resistant to chloramphenicol, ampicillin and

TABLE-I

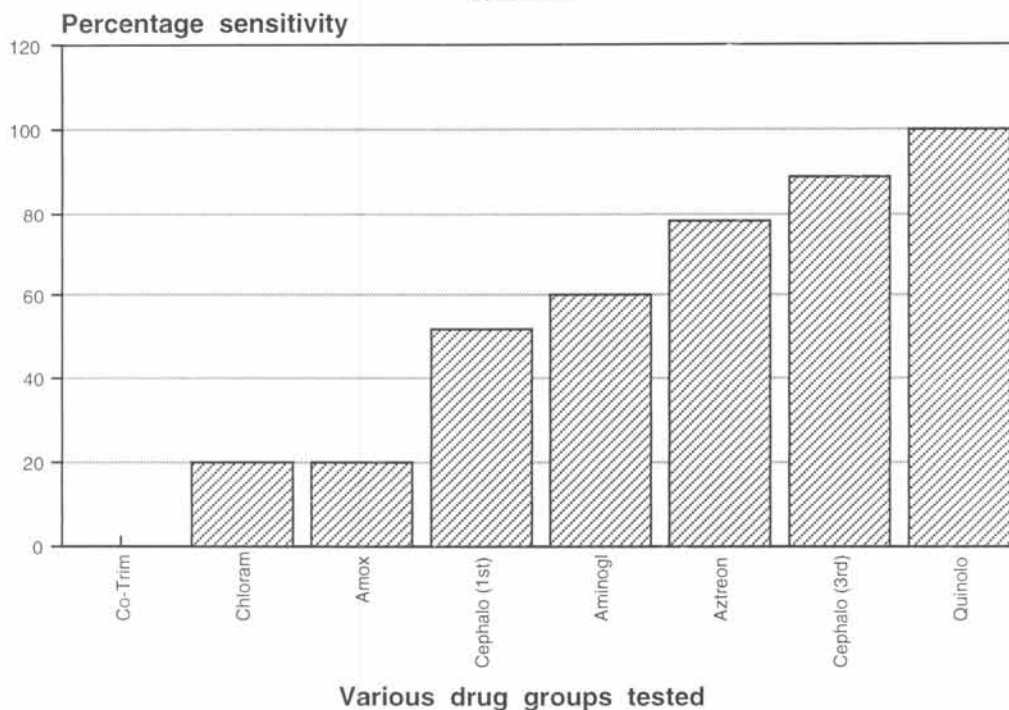
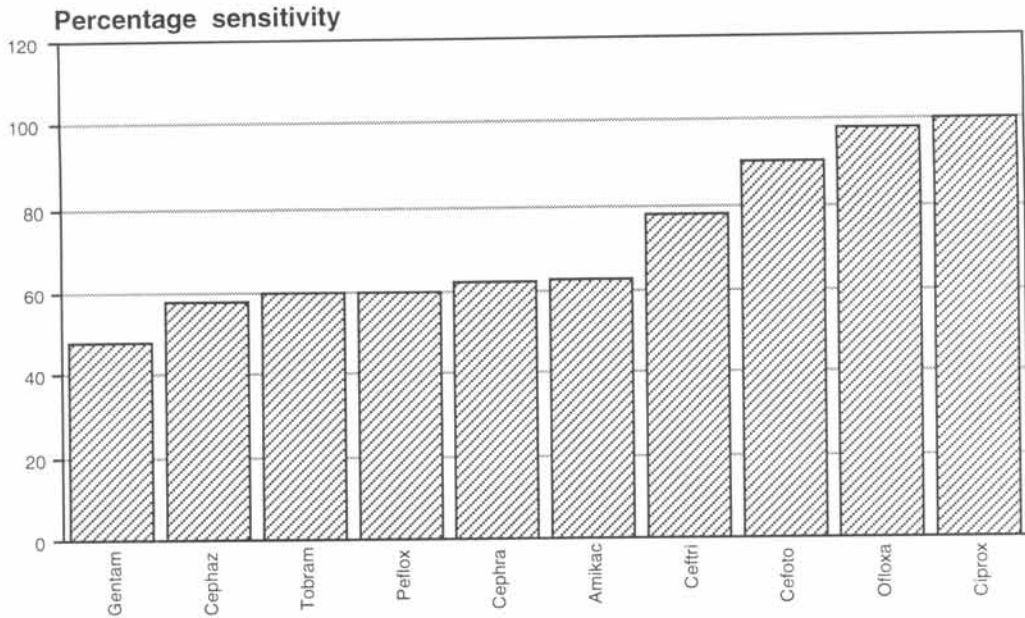


TABLE-II



Sensitivity to various drugs tested

SMZ-TMP have been reported from Pakistan, India, Vietnam, California, North Eastern United States and Spain. This resistance has increased in some series from 16% to 80% in India.^{5,6,8}

In our patients the invitro resistance of Salmonella to SMZ-TMO was 100% and that to chloramphenicol and amoxycillin 80%. Resistance to aminoglycosides was from 36 to 65%. However in our clinical practice none of the three patients responded to amoxycillin (0%) in spite of the positive culture and sensitivity report from the laboratory i.e. invitro study. A similar change was observed for chloramphenicol where the clinical outcome showed no improvement in 50% of the cases in spite of the recorded 20% sensitivity by the laboratory results.

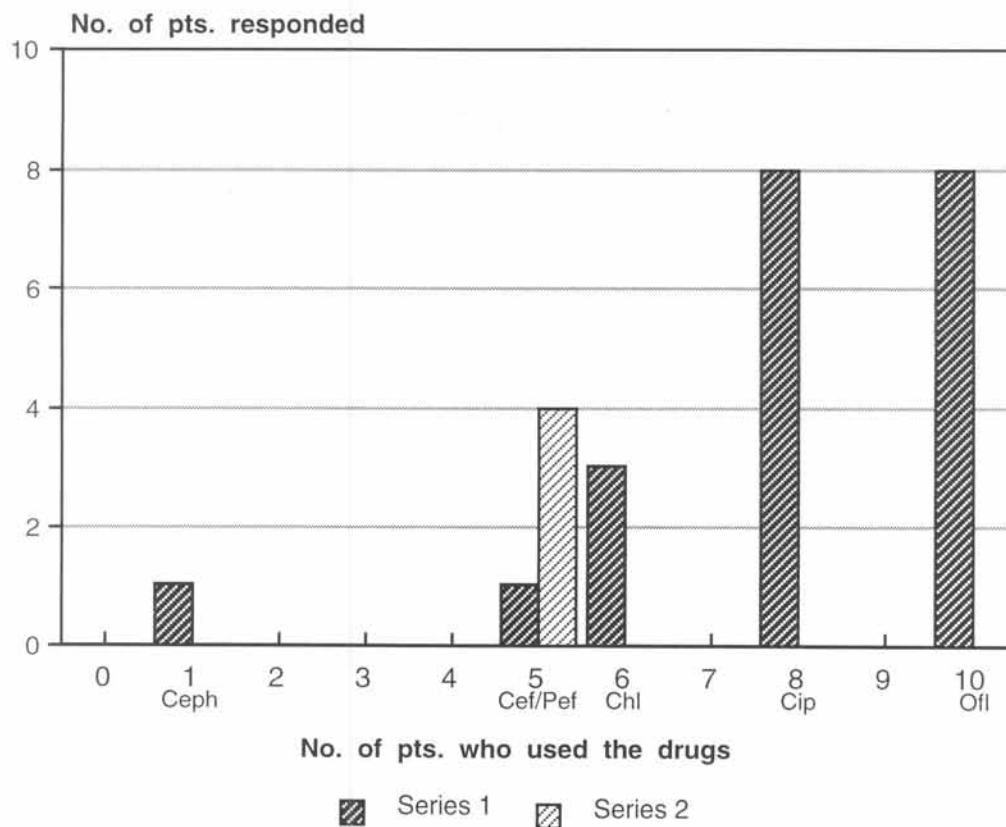
Resistance to these commonly used drugs mentioned above needs a change in therapeutic approach to typhoid fever. In this search cephalosporins have been studied and both cephaloridine and cephradine

were effectively used in small series of patients. Paradelis (1980) also found the multi-resistant strains very sensitive to cephradine and treated 10 such patients successfully with it.⁹ The slightly patients in our series in whom it was used responded well and the use of these drugs needs further clinical evaluation.

Third generation cephalosporins have good tissue penetration and prolonged antibacterial activity. Girgis (1993) reported cefixime to be a very effective oral drug.¹⁰ Cefomandal is successful only when it is used as a continuous infusion for 10-15 days. There are many randomized comparative studies of the efficacy of ceftriaxone.²

In our study the invitro sensitivity of 3rd generation cephalosporins was 96%. Cefotaxime (88%) was better than ceftriaxone (77%) in the patients tested. In clinical practice ceftriaxone was administered to 5 patients of whom only one (20%) responded. These results show the same discrepancies between in-vitro studies and

TABLE-III
CLINICAL RESPONSE OF PATIENTS TO VARIOUS DRUGS



clinical effectiveness. In spite of the cost of the drug, exceedingly large doses i.e. 2gm BD and the intravenous route which necessarily requires hospital admission, the results were disappointing.

Ciprofloxacin has excellent activity against the enterobacteriaceae, including the majority of organisms resistant to currently available penicillins, cephalosporins and aminoglycosides. The drug was 100% effective in clinical set up.^{5,11} Ofloxacin and fleroxacin were also 100% effective and the clinical cure rate with pefloxacin was above 90%.¹²⁻¹⁴ The minimum inhibitory concentration of rifloxacin against *S.typhi* is reported to be 4-16 times higher than the above mentioned quinolones.¹⁵

In our patients the in-vitro activity of quinolones was 100%. Ciprofloxacin was 100% effective as compared with ofloxacin (96%) and pefloxacin (60%). The clinical efficacy of ciprofloxacin was 100% in eight patients. However of all the 10, only 8 (80%) responded to ofloxacin. Clinical efficacy of pefloxacin was also 80%. As before, discrepancies persist between clinical responses and the laboratory data. In spite of 96% sensitivity to ofloxacin from the laboratory clinical improvement was observed in only 80%. However the results for ciprofloxacin remained consistent.

Salmonella has shown a gradual increase in its resistance over the years to the newly introduced quinolones, thus posing a major threat to the community and forcing

us to search for alternative means and antibiotics to combat this threat.¹⁶ In our series 76% of the patients were sensitive to Aztreonam. Farid and Girgis (1987) used this drug in eleven patients all of whom were cured.¹⁰ The oral monobactam e.g. Tigenim has been reported to inhibit 90% of strains of E-coli, Klebsiella and Salmonella. Carbapenems (biapenem, imipenem, meropenem) in-vitro inhibit 90% of isolates of enterobacteriaceae and Salmonella typhi. 1st use in children eradicated Salmonella spp.^{17,18} The in-vitro activity of azithromycin has been found to be very high against enteric pathogens including Salmonella typhi. Gradello Me (1993) has suggested its use in such infections.¹⁹ The results of the use of Timocillin in enteric fever were very promising in Thailand.²⁰ The new parenteral oxime-type cephalosporin, FK-037, inhibits 90% isolates of Salmonella typhi.²¹ The antibiotic susceptibility of Salmonella is enhanced many folds to cefuroxime, ciprofloxacin, chloramphenicol and rifampicin when lactoferrin is combined with these drugs.²²

As is obvious from the rising resistance to various antibiotics, nature is on the side of the bacteria. The dwindling list of drugs leads us to consider other means of control. Some of these may be as follows:

1. Education in the basic principles of hygiene.
2. The supply of clean drinking water to every household and frequent inspections by the water authorities.
3. Patient isolation and facilities for the disposal of excrement.
4. Immunization

Where sophisticated and expensive drugs have failed commonsense may succeed. An old English proverb says, "an ounce of prevention is better than a pound of cure." What the third world needs is better living conditions. It may be more

sensible to funnel funds into improving these rather than develop new drugs to combat established disease.

REFERENCES

1. Edwards CRW, Bouchier IAD, Hosle HC, Chilvers ER. Davidson's principles and practice of Medicine. 17th ed. Edinburgh Churchill Livingstone. 1995; 122.
2. Herzog C, Geddes AM. Chloramphenicol in the treatment of Enteric fever. Trans Soc Trop Med Hyg. 1982; 76(6): 848.
3. Smego RA, Zaidi AK, Mohammad Z, et al. Multiply resistant Salmonella and Shigella Isolates. APMIS Suppl 1988; 3: 65.
4. Chakrovorty B, Jain N, Gupta B, et al. Chloramphenicol Resistant Enteric Fever. J Ind Med Assoc. 1993; 91(1): 10.
5. Karamat KA, Mehmood AA, Iftikhar A. Multidrug resistant Salmonella typhi and Ciprofloxacin Ciproxin trial report. Bayer Monograph 22.
6. Li-Q. Surveillance on vill phage typing and antimicrobial susceptibilities of S.typhi strains in Guangdong province Chung Hua Liu Hsing Ping Hsueh Tsa Chih. 1992; 12: 288.
7. Solodovnikov LUP, Novoselova ELU. Sensitivity to antibacterial drugs and phage type pattern of Salmonella typhi isolate from patients in 1990 Antibiot Khimioter. 1993; 38(8): 39.
8. Rodrigues C, Metha A, Mehtar S, et al. Chloramphenicol resistance in Salmonella typhi; Report from Bombay J Assoc Physicians India 1992; 40(11): 729.
9. Paradelis AG, Edipides T, Zurukzoglov W. Recent advance in Salmonellosis treatment based on in-vitro susceptibilities Methods Find Exp Clin Pharmacol. 1980; 2(6): 313.
10. Farid Z, Girgis NL, et al. Aztreonam in the treatment of enteric fevers. Annals of Tropical Medicine and Parasitology 1987; 9: 725.
11. Lasserre R, Sangalang RP, Santiags L. Three days treatment of Enteric fever with two different doses of Ceftriaxone, com-

- pared to 14 days therapy with Chloramphenicol a randomized Trial. *J Antimicrobial Chemotherapy*. 1991; 28: 765.
12. Arnold K, et al. Randomized comparative study of Fleroxacin and Chloramphenicol in Typhoid fever: *Am J Med* 1993; 94 (suppl 3A) 195.
 13. Sabbour MS, Osmo LM. Experience with Ofloxacin in Enteric fever. *J Chemother* 1990; 2(2): 113.
 14. Shujaat H, Khan A, Ahmad M. Management of Typhoid fever with Pefloxacin An Opemn, Single drug non-comparative trial *Pak Armed Forces Med J* 1989; 44(2): 10.
 15. Qadri SM, Ayub A, Veno Y, Saldin H. Susceptibility of *Salmonella typhi* and *Brucella melitensis* to the new fluorouinolone rufloxacin. *Chemotherapy* 1993; 39(5): 311.
 16. Hafiz S, Khan SW, Sharif R, et al. Epidemiology of Salmonellosis and its sensitivity in Karachi. *J. Pak Med Assoc.* 1993; 43(9): 178.
 17. Clark AM, ZemcovSJ. Comparative in-vitro activity of Biapenem, a new Carbapenem antibiotic. *Eur J Clin Microbiol Dis* 1993; 12(5): 337.
 18. Tajima T, Kobayashi M, Heta M, et al. Pharmacokinetics, Bacteriological and clinical studies on SY 5555 in children. *Jpn J Antibiotic* 1995; 48(1): 31.
 19. Gordilo ME, Sigh KV, Morray BE. In-vitro activity of Azithromycin against bacterial enteric pathogens. *Antimicrob-Agents-Chemother* 1993; 37(5): 1203.
 20. Tanphachitra D, Kanjanapanjapal S, Srimuang S. Use of timocillin in typhoid fever, hepatobiliary diseases and other infections. *Drugs* 1985; 29(5): 201.
 21. Clark AM, Secov SJ, Hubinettee MM. Comparative in-vitro activity of, Fk-037, a new Cephalosporin antibiotic. *Diagn Microbiol Infect Dis.* 1994; 20(1): 27.
 22. Daidu AS, Arnold RR. Lactoferrin interaction with *Salmonella* patentiate antibiotic susceptibility in vitro. *Diagn Microbial Infect Dis* 1994; 20(2): 69.